

MALDI-TOF Mass Spectral Characterization of Polymers Containing an Azide Group: Evidence of Metastable Ions

Yejia Li, Jessica N. Hoskins, Subramanya G. Sreerama, and Scott M. Grayson*

Department of Chemistry, Tulane University, New Orleans, Louisiana 70118 Received April 2, 2010; Revised Manuscript Received June 11, 2010

Introduction

The copper-catalyzed Huisgen 1,3-dipolar cycloaddition "click" reaction, ¹ has become an invaluable coupling reaction due to its uncommon combination of extremely high coupling yields and broad functional group tolerance. This versatility has been demonstrated in numerous applications² including the labeling of biological molecules (DNA, proteins, cell surfaces, etc.) both in vitro and in vivo, ^{3,4} synthesis of biomedical materials, ⁵ the cross-linking self-assembled nanostructures, ⁶ and the functionalization of two-dimensional surfaces. ⁷

In the field of macromolecular synthesis, the broad utility of click coupling has been proven through the assembly of an exceptional range of complex macromolecular architectures, including diblock copolymers, triblock copolymers, star polymers, triblock copolymers, tar polymers, are polymers, are polymers, are polymers, dendriners, below the dendrimers, for dendriners below to copolymers, below to polymers, for cyclic polymers, for cyclic block copolymers, below to polymers, and even dendrimers with polymeric repeat units. As a consequence, characterization of well-defined azide-functionalized macromolecules has become an increasingly important aspect of polymer chemistry.

One of the most valuable tools for the characterization of polymers is matrix-assisted laser desorption ionization time-of-flight mass spectrometry (MALDI-TOF MS), which provides a means to determine the $M_{\rm n}$ and PDI of a polymer sample, as well as to accurately calculate the repeat unit and end group masses. In most traditional end group identification techniques, such as NMR or IR, the end group signal becomes vanishingly small relative to that of the repeat units as molecular weight increases. However, in a polymer's mass spectrum, each signal contains information about that n-mer's end groups, and therefore the mass of the end groups can be easily calculated as long as the mass of the individual signals can be accurately determined. ²¹

Despite the importance of monoazide functionalized polymers, their reported MALDI-TOF reflectron mass spectra have frequently been misinterpretated; a result of the unusual susceptibility of the azide functionality to fragment during matrix-assisted laser desorption ionization. Mass spectra obtained in our laboratories and elsewhere suggest that fragmentation of the azide functionality via expulsion of N_2 gives rise to ions 28 mass units less than the mass of the *n*-mer. ²² However, a distribution is observed in addition to the [M-28] ions in reflector mass spectra, which has a mass loss less than 28 mass units, thus exhibiting the characteristics of metastable ions. This later distribution can be misinterpreted if one is not familiar with the metastable ion formation process in mass spectrometry. In order to confirm the proposed metastable ion formation in reflector mass spectra of

Scheme 1. Chemical Structures of the PCL, Cyclic PCL, PEG, and PS Monoazide Polymers Used in This Study (PMDETA = N,N,N',N'', N''-Pentamethyldiethylene Triamine)

polymer azides, a variety of different monoazide functionalized polymers were examined, including polycaprolactone (PCL), poly(ethylene glycol) (PEG), and polystyrene (PS) (Scheme 1).

Results and Discussion

Metastable ions are formed in MALDI–TOF mass spectrometry when a parent ion generated during laser desorption fragments at some point during the flight path in the field free region between the ion source and detector. For the azide functional group, facile fragmentation has precedence in small molecule analogues, including the thermal fragmentation of phenyl azides by expulsion of N_2 to generate reactive nitrene intermediates, as well as analogous fragmentation pathways in the mass spectra of low molecular weight aryl and aliphatic azides.

In order to verify postsource metastable ion formation occurs for polymeric azides, highly precise end group mass determinations must be carried out. In this study, the accurate determination of end group masses within 0.01 Da (in the case of PEG) was made possible by a combination of high resolving power instrumentation (Bruker Autoflex III) and a rigorous eight point calibration across the molecular weight range (using monodisperse dendrimer standards).²⁷

^{*}To whom correspondence should be addressed. E-mail: sgrayson@tulane.edu.

6226

Figure 1. Comparison of MALDI-TOF mass spectra obtained for polycaprolactone ($M_{\rm n}=2040~{\rm PDI}=1.04$) in (a) reflector mode, and (b) in linear mode and for poly(ethylene glycol) ($M_{\rm n}=2193~{\rm PDI}=1.01$) in (c) reflector mode, and (d) in linear mode.

As seen in Figure 1, monoazide functionalized PCL and PEG exhibit three distinct distributions in their mass spectra when analyzed using reflector mode: the parent ion $([M + Na]^{+})$ in blue), the in-source expulsion of N_2 ([M – 28 + Na]⁺ in green) and the proposed postsource expulsion of N₂ (metastable ions in red). The first observation that provides strong evidence for the metastable nature of this third distribution involves its disappearance from the spectra when linear mode is used instead of reflector mode. In linear mode, postsource metastable ion formation in the field free region will produce smaller neutral and ionic fragments with the same velocity, and hence time-of-flight, as the parent ion. As a result, such fragmentation will not be apparent in mass spectra acquired in linear mode. In reflector mode, however, ions are decelerated partway through their flight path and then reaccelerated in a different direction by a "mirror" toward the second detector, in order to provide enhanced resolving power. Any postsource metastable ions formed before arriving at the mirror will undergo a different deceleration, reacceleration than their parent ion. As a result, the time-offlight, and thus the mass for those metastable ions (the red series) will be some intermediate value between that of the parent mass (the blue series) and the in-source fragment mass (the green series). It should be noted that the intensity of the metastable signal increases with laser power (Figure S1, Supporting Information). As a result, polymers that require higher laser power for ionization will exhibit more intense metastable signals (for example, polystyrene in Figure 2c).

A further indication of metastable ion formation in polymer azides is the nonuniform, noninteger mass offset of the third (red) distribution relative to parent azide distribution. If this distribution were the result of a degradation product or a different mode of ionization, the offset should be uniform across the entire

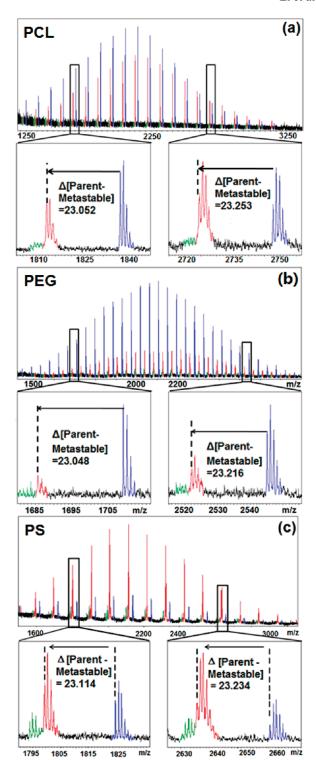


Figure 2. Comparison of reflector mode MALDI-TOF mass spectra for (a) poly(ε -caprolactone) ($M_{\rm n}=2040\,{\rm PDI}=1.04$) (b) poly(ethylene glycol) ($M_{\rm n}=2190\,{\rm PDI}=1.01$) and (c) polystyrene ($M_{\rm n}=2160\,{\rm PDI}=1.03$), highlighting the difference between the parent ion and the metastable ion at both the low and high extremes of the mass distributions

molecular weight range. In the case of PEG-N₃, the parent (blue distribution) and in-source fragment (green distribution) ions correspond closely with the theoretical masses (each observed parent ion within 0.01 Da of the theoretical value). However, the metastable (red distribution) ions exhibit a nonuniform mass offset that increases at higher molecular weights (Tables S1–S3 in the Supporting Information). For example, for the PEG 13-mer

near 652 Da, the mass offset relative to its parent ion is approximately 22.67 Da, however, near 2633, the analogous mass offset is greater than 23.25 Da (Figure 2). This discrepancy of nearly 0.60 Da is almost 2 orders of magnitude greater than the observed error for the calibrated parent ion data over the same mass range. A similar trend is observed for the other two polymers investigated, (though the signal of the other backbones was more obscured by background noise due to inefficient ionization) where increasing molecular weight of the parent ion results in an increased offset in the metastable ion (Figure 3). This noninteger mass results from postsource metastable ion formation, since the ion mass changes after its original acceleration from the source and before its deceleration/reacceleration by the mirror. Because the lighter metastable ions lose a greater percentage of their mass during the azide fragmentation, they are affected more strongly by the reflector voltage, and exhibit an observed mass closer to the parent ion, while heavier ions exhibit an observed mass closer to the in-source fragment ion. This nonuniform metastable mass shift has been modeled elsewhere, ²⁸ and the previously reported trends agree with the data observed in this study.

Additional confirmation of the metastable nature of the azide end groups is the disappearance of both the in-source and postsource fragmentation ions in the polymer mass spectra after their "click" coupling, because the product triazole is aromatic and therefore is no longer susceptible to fragmentation. A particularly clear example of this is after the intramolecular cyclization of polycaprolactone with complementary azide and alkyne end groups on opposite ends of the polymer. ²⁹ Because the Huisgen 1,3-dipolar cycloaddition is an intramolecular pericyclic reaction, there is no loss or gain in the molecular mass; however,

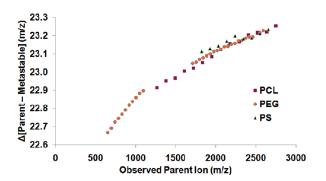


Figure 3. Graph comparing the mass discrepancies between the metastable signals and the parent signals for three monoazide functionalized polymers: PCL (purple), PEG (orange), and PS (black).

the fragmentation of the parent ion is no longer plausible in the triazole product and as a result the signal corresponding to the parent ions were retained while those corresponding to the azide fragments are lost (Figure 4).³⁰

Conclusion

In conclusion, the mass spectra of monoazide functionalized polymers have been investigated in detail to verify the prevalence of fragment ions resulting from a loss of N_2 via both in-source and postsource metastable ion formation. The postsource metastable ions are identified by a nonuniform, noninteger mass offset relative to the parent ion, and can be most easily confirmed by their disappearance using linear mode detection. With the increasing importance of macromolecular azides in polymer research, the authors hope this study will assist others examining their mass spectra by facilitating the identification and interpretation of the metastable signals.

Acknowledgment. The authors thank Michael A. Grayson and Paul Kowalski for helpful discussions, the NSF for providing MALDI—TOF MS instrumentation (MRI 0619770), as well as Tulane University, the National Institutes of Health (5R01EB-6493), the Louisiana Board of Regents (LEQSF(2006-09)-RD-A-29), the donors of the Petroleum Research Fund (47108-G7), administered by the American Chemical Society, and the National Science Foundation (NSF-DMR 0844662 (ARRA)) for financial support.

Supporting Information Available: Text giving experimental synthetic and analytical procedures, tables of MALDI-TOF data, schemes showing the reactions used, and a figure showing examination of metastable signal intensity with respect to increasing laser power. This material is available free of charge via the Internet at http://pubs.acs.org.

References and Notes

- Huisgen, R. In 1,3-Dipolar Cycloaddition Chemistry; Padwa, A., Ed.; Wiley: New York, 1984; Kolb, H. C.; Finn, M. G.; Sharpless, K. B. Angew. Chem., Int. Ed. 2001, 40, 2004. Tornøe, C. W.; Christensen, C.; Meldal, M. J. Org. Chem. 2002, 67, 3057–3064.
- (2) Meldal, M.; Tornøe, C. W. Chem. Rev. 2008, 108, 2952-3015.
- Amblard, F.; Cho, J. H.; Schinazi, R. F. Chem. Rev. 2009, 109, 4207–4220.
- (4) Best, M. D. Biochemistry 2009, 48, 6571-6584.
- (5) van Dijk, M.; Rijkers, D. T. S.; Liskamp, R. M. J.; van Nostrum, C. F.; Hennink, W. E. *Bioconjugate Chem.* 2009, 20, 2001–2016. Lutz, J.; Zarafshani, Z. Adv. Drug Delivery Rev. 2008, 60, 958–970.
- (6) Joralemon, M. J.; O'Reilly, R. K.; Hawker, C. J.; Wooley, K. L. J. Am. Chem. Soc. 2005, 127, 16892–16899.

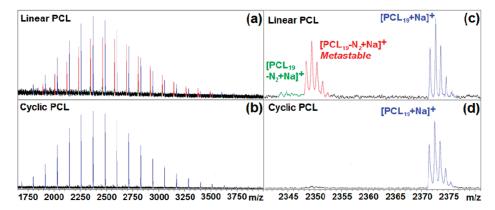


Figure 4. Comparison of MALDI-TOF mass spectra obtained for (a) linear poly(ε -caprolactone) ($M_n = 2440$ PDI = 1.02) and (b) cyclic poly(ε -caprolactone) ($M_n = 2400$ PDI = 1.02). Insets c and d correspond to linear and cyclic poly(ε -caprolactone), respectively, where the expanded region between 2340 and 2380 m/z, highlights the disappearance of the (red) postsource fragment ions and the (green) in-source fragment ions as a result of the aromatic stabilization of the triazole product.

- Bryan, M. C.; Fazio, F.; Lee, H. K.; Huang, C. Y.; Chang, A.; Best, M. D.; Calarese, D. A.; Blixt, O.; Paulson, J. C.; Burton, D.; Wilson, I. A.; Wong, C. H. J. Am. Chem. Soc. 2004, 126, 8640–8641. Collman, J. P.; Devaraj, N. K.; Chidsey, C. E. D. Langmuir 2004, 20, 1051–1053. Meng, J. C.; Siuzdak, G.; Finn, M. G. Chem. Commun. 2004, 10, 2108–2109. Sun, X. L.; Stabler, C. L.; Cazalis, C. S.; Chaikof, E. L. Bioconjugate Chem. 2006, 17, 52–57. Santoyo-Gonzalez, F.; Hernandez-Mateo, F. Chem. Soc. Rev. 2009, 38, 3449–3462.
- (8) Fournier, D.; Hoogenboom, R.; Schubert, U. S. Chem. Soc. Rev. 2007, 36, 1369–1380. Lutz, J. F. Angew. Chem., Int. Ed. 2007, 46, 1018–1025.
- (9) Opsteen, J. A.; van Hest, J. C. M. Chem. Commun. 2005, 57–59. Quémener, D.; Davis, T. P.; Barner-Kowollik, C.; Stenzel, M. H. Chem. Commun. 2006, 5051–5053.
- (10) Durmaz, H.; Dag, A.; Altintas, O.; Erdogan, T.; Hizal, G.; Tunca, U. *Macromolecules* 2007, 40, 191–198.
- (11) Gao, H.; Matyjaszewski, K. Macromolecules 2006, 39, 4960-4965.
- (12) Whittaker, M. R.; Urbani, C. N.; Monteiro, M. J. J. Am. Chem. Soc. 2006, 128, 11360–11361.
- (13) Gao, H.; Matyjaszewski, K. J. Am. Chem. Soc. **2007**, 129, 6633–6639.
- (14) Scheel, A. J.; Komber, H.; Voits, B. I. Macromol. Rapid Commun. 2004, 25, 1175–1180.
- (15) Wu, P.; Feldman, A. K.; Nugent, A. K.; Hawker, C. J.; Scheel, A.; Voit, B.; Pyun, J.; Fréchet, J. M. J.; Sharpless, K. B.; Fokin, V. V. Angew. Chem., Int. Ed. 2004, 43, 3928–3982. Joralemon, M. J.; O'Reilly, R. K.; Matson, J. B.; Nugent, A. K.; Hawker, C. J.; Wooley, K. L. Macromolecules 2005, 38, 3663–3678.
- (16) Helms, B.; Mynar, J. L.; Hawker, C. J.; Fréchet, J. M. J. J. Am. Chem. Soc. 2004, 126, 15020–15021.
- (17) Laurent, B. A.; Grayson, S.M. J. Am. Chem. Soc. 2006, 128, 4238– 4239
- (18) Eugene, D. M.; Grayson, S. M. Macromolecules 2008, 41, 5082-5084.
- (19) Loethen, S.; Ooya, T.; Choi, H. S.; Yui, N.; Thompson, D. H. Biomacromolecules 2006, 7, 2501–2506.
- (20) Urbani, C. N.; Bell, C. A.; Whittaker, M. R.; Monteiro, M. J. Macromolecules 2008, 41, 1057–1060.
- (21) The use of MALDI-TOF mass spectrometry for polymer end group analysis has evolved alongside instrument development, a brief review and leading references can be found in: Li, Y.; Hoskins, J. N.; Sreerama, S. G.; Grayson, M. A.; Grayson, S. M.

- J. Mass Spectrom. 2010, 45, 587–611. In addition, a number of the foundational papers include the following: Danis, P. O.; Karr, D. E.; Mayer, F.; Holle, A.; Watson, C. H. Org. Mass Spectrom. 1992, 27, 843–846. Maloney, D. R.; Hunt, K. H.; Lloyd, P. M.; Muir, A. V. G.; Richards, S. N.; Derrick, P. J.; Haddleton, D. M. Chem. Commun. 1995, 561–562. Montaudo, G.; Montaudo, M. S.; Puglisis, C.; Samperi, F. Macromolecules 1995, 28, 4562–4569. Danis, P. O.; Karr, D. E.; Simonsick, W. J., Jr.; Wu, D. T. Macromolecules 1995, 28, 1229–1232. Zammit, M. D.; Davis, T. P.; Haddleton, D. M.; Suddaby, K. G. Macromolecules 1997, 30, 1915–1920. Williams, J. B.; Gusev, A. I.; Hercules, D. M. Macromolecules 1997, 30, 3781–3787.
- (22) For polystyrene: Matyjaszewski, K.; Nakagawa, Y.; Gaynor, S. G. Macromol. Rapid Commun. 1997, 18, 1057–1066. Coessens, V.; Matyjaszewski, K. J. Macromol. Sci., Part A: Pure Appl. Chem. 1999, A36, 667–679. Lutz, J. F.; Borner, H. G.; Weichenhan, K. Macromol. Rapid Commun. 2005, 26, 514–518. Guillaneuf, Y.; Dufils, P.-E.; Autissier, L.; Rollet, M.; Gigmes, D.; Bertin, D. Macromolecules 2010, 43, 91–100. For poly(ethylene glycol): Raynaud, J.; Absalon, C.; Gnanou, Y. J. Am. Chem. Soc. 2009, 131, 3201–3209.
- (23) Spengler, B.; Kirsch, D.; Kaufmann, R.; Cotter, R. J. Rapid Commun. Mass Spectrom. 1991, 5, 198–202. Kaufmann, R.; Chaurand, P.; Kirsch, D.; Spengler, B. Rapid Commun. Mass Spectrom. 1996, 10, 1199–1208. Spengler, B. J. Mass Spectrom. 1997, 32, 1019–1036.
- (24) Bertho, A. Chem. Ber. 1924, 57, 1138-1142.
- (25) Crow, W. D.; Wentrup, C. Tetrahedron Lett. 1967, 44, 4379–4384. Abramovitch, R. A.; Kyba, E. P.; Scriven, E. F. V. J. Org. Chem. 1971, 36, 3796–3803.
- (26) Olivera, A. M.; Barros, M. T.; Martins, A. M.; Cabral, M. A. R.; Dias, A. A.; Costa, M. L.; Cabral, M. H.; Moutinho, A. M. C.; Jennings, K. R. Rapid Commun. Mass Spectrom. 1999, 13, 559–561.
- (27) Grayson, S. M. US Patent 2010/023087 pending.
- (28) Harvey, D. J.; Hunter, A. P.; Bateman, R. H.; Brown, J.; Critchley, G. Int. J. Mass. Spectrom. 1999, 188, 131–146. Stairs, J. R.; Dermota, T. E.; Wisniewski, E. S.; Castleman, A. W., Jr. Int. J. Mass. Spectrom. 2002, 213, 81–89. Shard, A. G.; Gilmore, I. S. Int. J. Mass. Spectrom. 2008, 269, 85–94.
- (29) Hoskins, J. N.; Grayson, S. M. Macromolecules 2009, 43, 6406–6413
- (30) The very low remaining signal at 2349 Da is likely caused by a trace amount of unreacted linear precursor.